# Multi-Institutional Hematopathology Interesting Case Conference

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## **Clinical History**

68-year-old male presented with skin lesions

#### Past Medical History

50 pack-year smoking history, thoracic aortic aneurysm, chronic kidney disease stage 3

#### Family history:

- Father passed away from leukemia in 1975.
- Brother treated for leukemia.

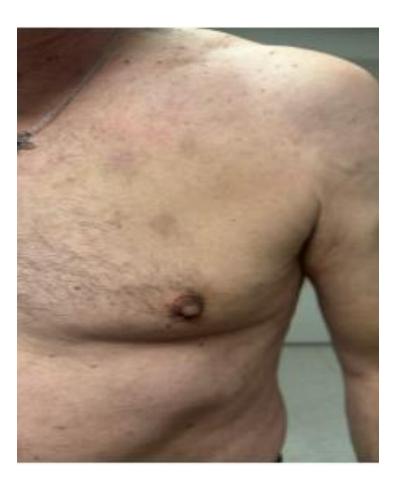
#### Physical Exam:

- Violaceous-pink raised painless skin lesions Upper extremities and torso with an intermittent burning sensation
- No palpable adenopathy
- No palpable hepatosplenomegaly

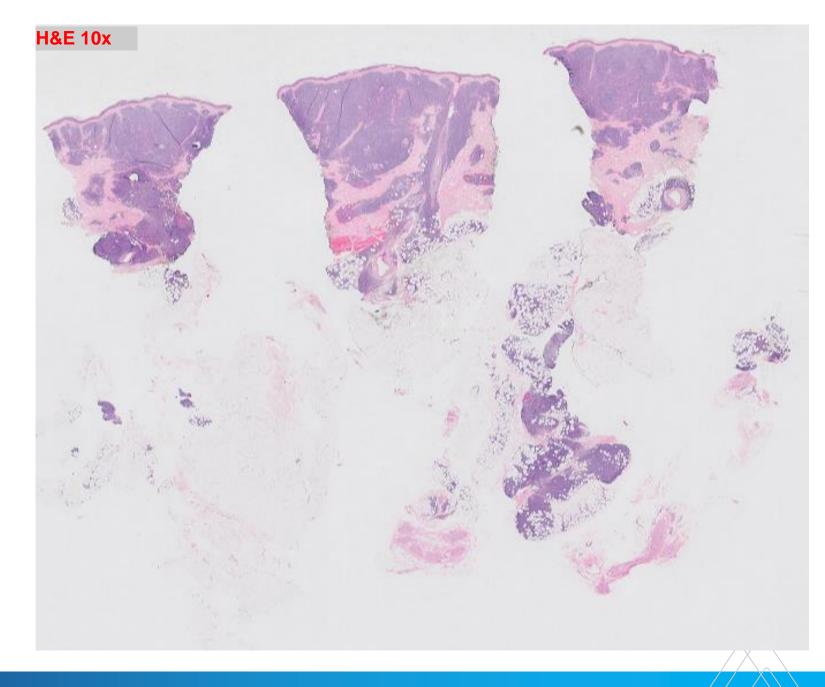
Underwent skin punch biopsy of right distal forearm

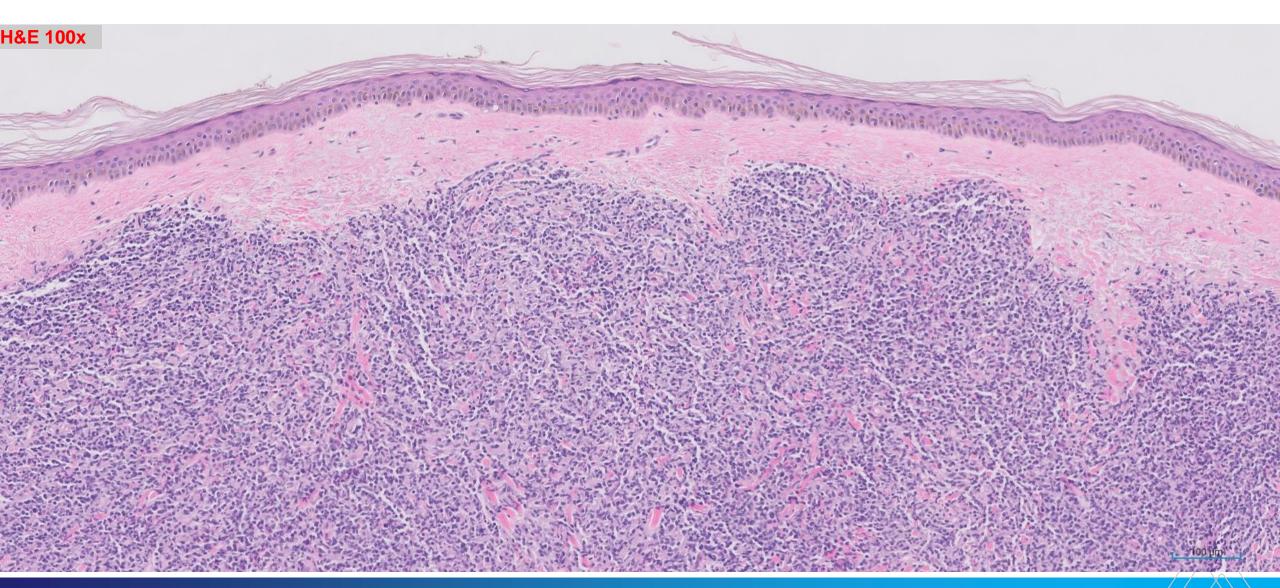
### **Clinical Photos**

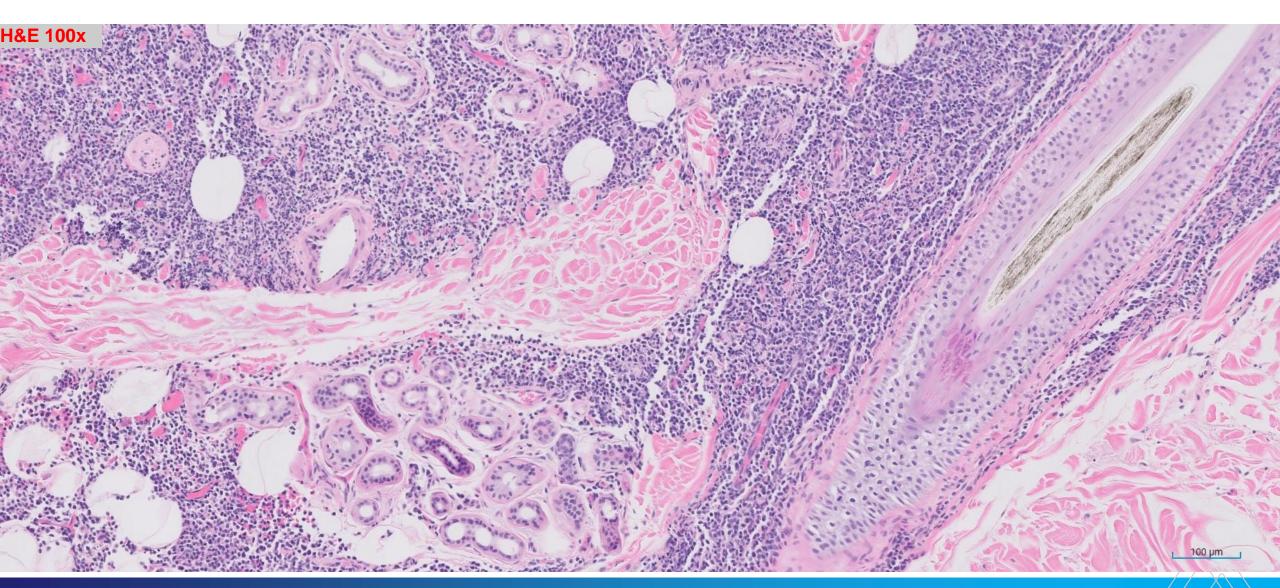


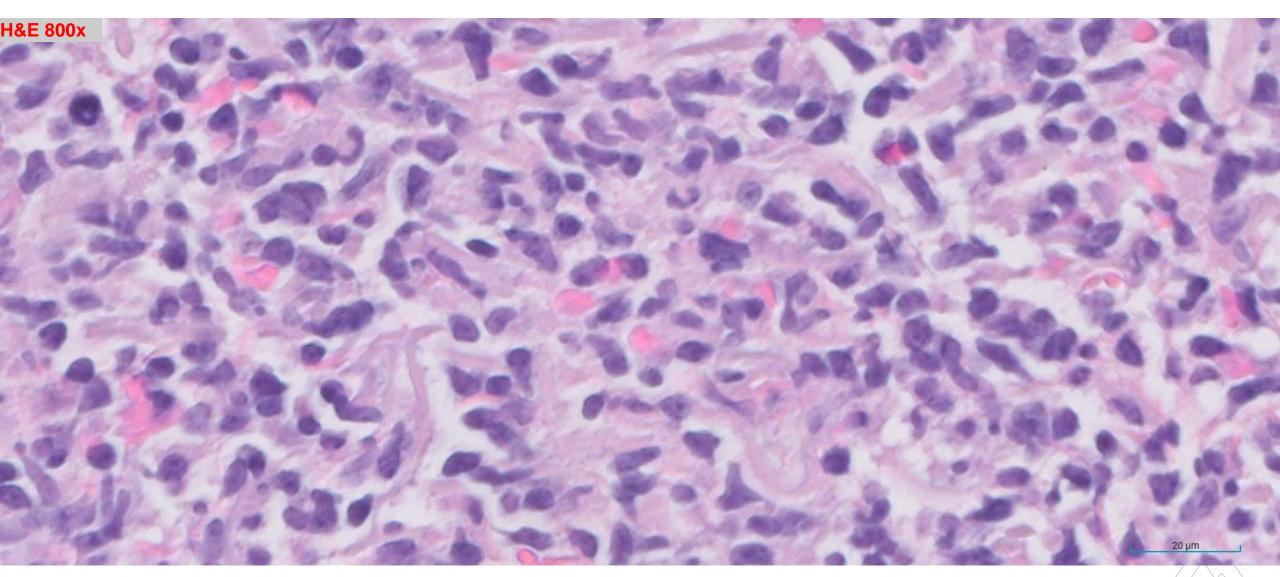


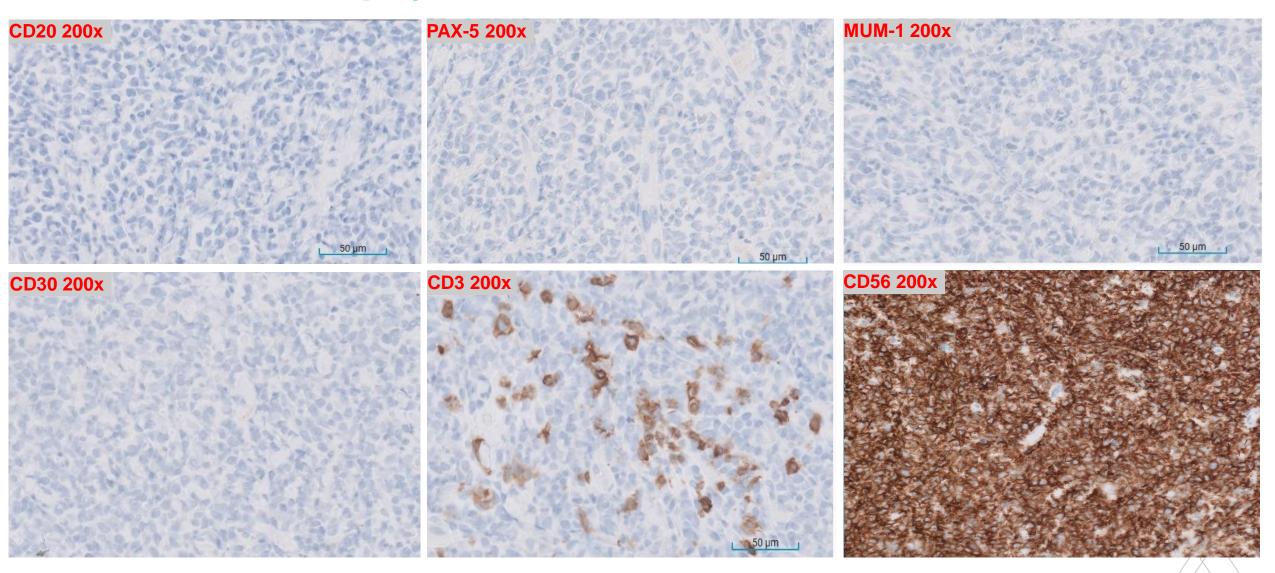












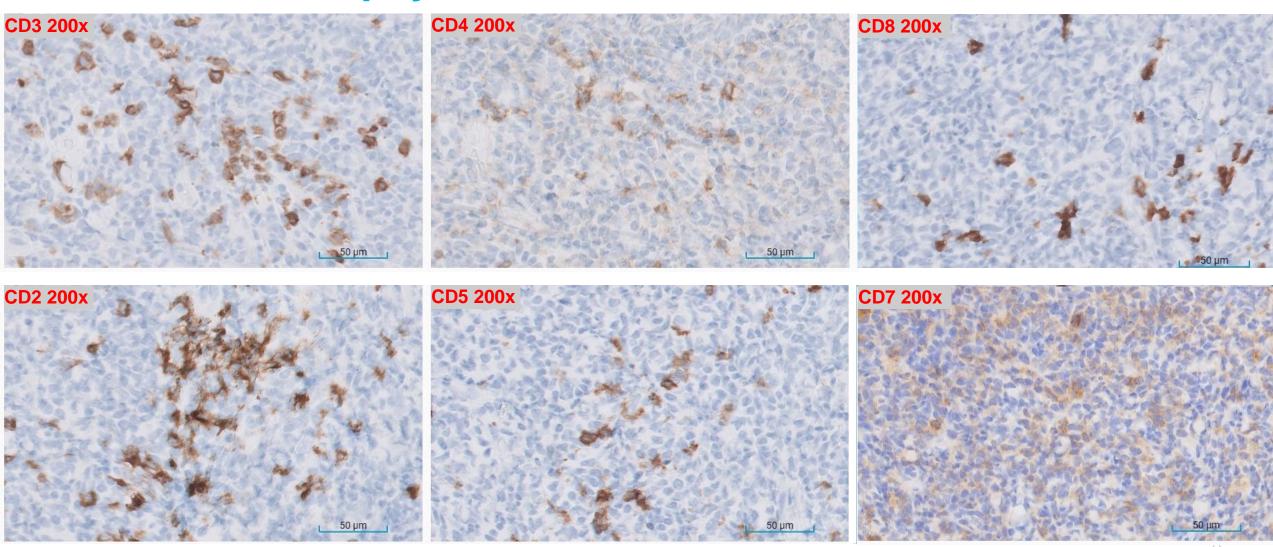
# Differential Diagnosis?

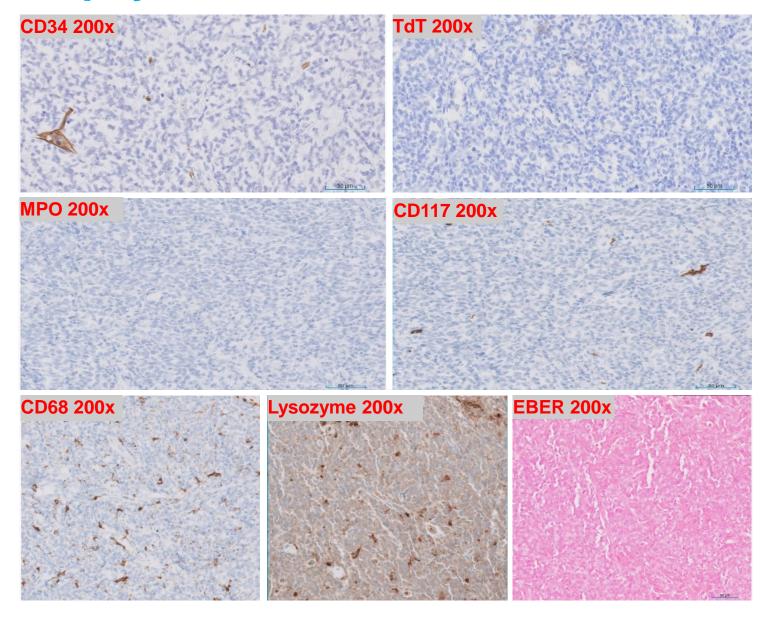
**NK/T** cell lymphoma

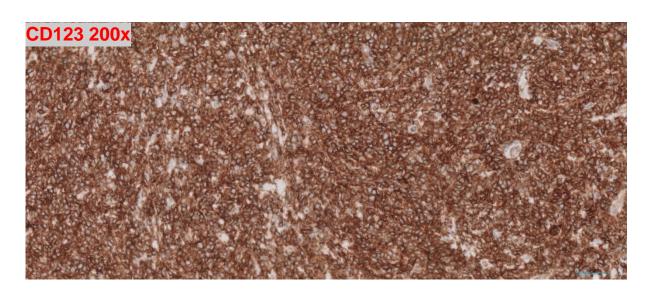
Leukemia cutis

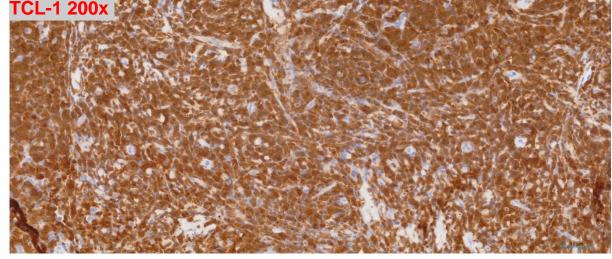
Histiocytic sarcoma

Blastic plasmacytoid dendritic cell neoplasm









## **Summary of Immunohistochemistry Results**

Samulary of the		moti y i toodito
Mar	Results	
Immaturity	CD34	Negative
	TDT	Negative
B cell and plasma cells	CD20	Negative
	PAX5	Negative
	MUM1	Negative
	CD2	Negative
	CD3	Negative
T cell	CD4	Negative
і сен	CD5	Negative
	CD7	Partial weak
	CD8	Negative
NK cell	CD56	STRONG
Myeloid	MPO	Negative
	Lysozyme	Negative
Monocytic	CD68	Negative
Dendritic	CD123	STRONG
	TCL1	STRONG
	EBER	Negative

#### **DIAGNOSIS:**

RIGHT DISTAL FOREARM, PUNCH BIOPSY-

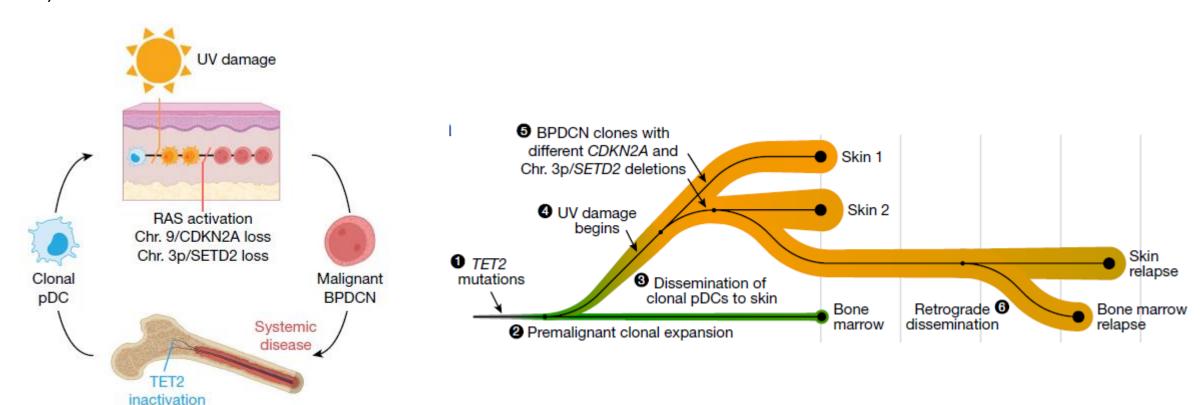
- BLASTIC PLASMACYTOID DENDRITIC CELL NEOPLASM

### Blastic plasmacytoid dendritic cell neoplasm

- Consists of immature cells with plasmacytoid dendritic cell (pDC) differentiation
- pDCs
  - Secrete interferon and act as antigen presenters
- Clinical features
  - Median age ~67 years
  - Male dominance
  - Skin lesions varying in size and color
- Genetic features
  - Proliferation and hematopoietic differentiation (ETV6 loss) (Tang, Zhenya, et al. Leukemia Research 2018)
  - Epigenetic regulation (TET2 and ASXL1) (Menezes J, et al. Leukemia 2014)
  - RNA splicing (e.g. SRSF2) (Brunetti et al, Leukemia. 2017)

#### **UV Radiation and BPDCN**

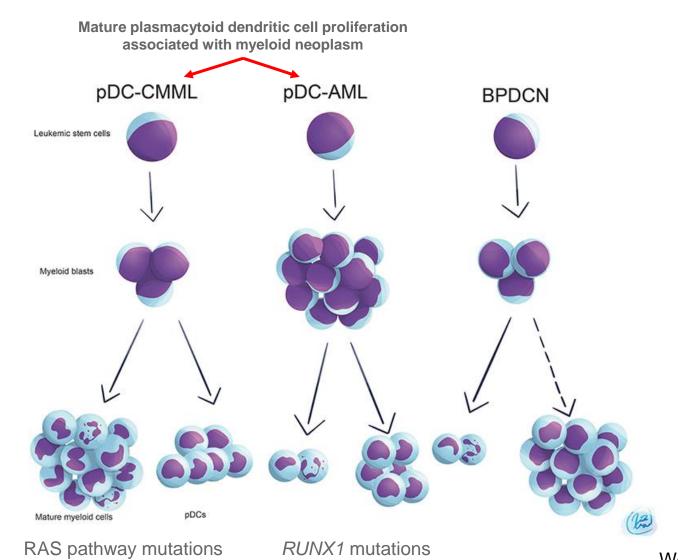
One study found that plasmacytoid dendritic cells derived from clonal BM progenitors seed the skin and subsequently transform to BPDCN in association with UV damage in sun-exposed sites (Griffin et al. *Nature*. 2023)



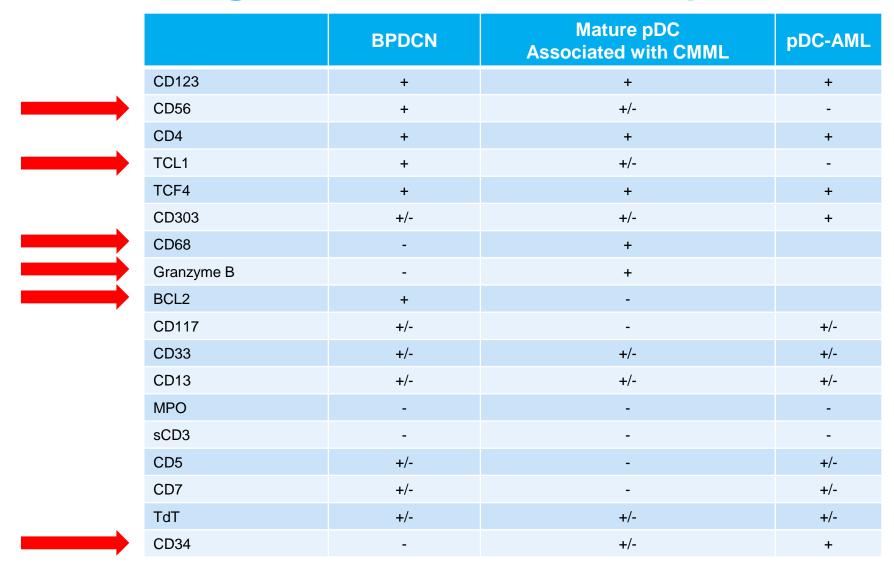
### Diagnostic criteria for blastic plasmacytoid dendritic cell neoplasm

World Health Organization 5th edition	International Consensus Classification 2022	
Expected positive markers	At least two of the following expressed	
pDC markers:	TCF4 CD422 CD202 TCF4 CD204	
CD123	TCF4, CD123, CD303, TCL1, CD304	
TCF4	Also typically expressed	
TCL1		
CD303	CD4	
CD304	0D-1	
Other markers:	CD56	
CD4		
CD56	Exclusionary markers	
Expected negative markers		
CD3	CD3	
CD14	CD19	
CD19	0510	
CD34	CD20	
Lysozyme	CD14	
Myeloperoxidase	CD14	
Immunophenotypic diagnostic criteria	Lysozyme	
Expression of CD123 and one other pDC marker in addition to CD4 and/or CD56		
OR	Myeloperoxidase	
Expression of any three pDC markers and absent expression of all expected negative markers	CD34	

# Other proliferations of pDCs



# Differentiating BPDCN from mature pDCs and pDC-AML



Wang, Wei, et al, Cancers (2022)

Vitte, Franck, et al. *The American Journal of Surgical Pathology* (2012)

# **CD4-negative BPDCN**

AJCP / ORIGINAL ARTICLE

# CD4-Negative Variant of Cutaneous Blastic Plasmacytoid Dendritic Cell Neoplasm With a Novel *PBRM1* Mutation in an 11-Year-Old Girl

Nuri Yigit, MD, <sup>1,3</sup> Luisa Fernanda Suarez, <sup>1</sup> Lisa Giulino Roth, MD, <sup>2</sup> Attilio Orazi, MD, <sup>1</sup> and Wayne Tam, MD, PhD<sup>1</sup>

From the <sup>1</sup>Division of Hematopathology, Department of Pathology and Laboratory Medicine, and <sup>2</sup>Department of Pediatrics, Weill Cornell Medical College, New York, NY; and <sup>3</sup>Department of Pathology, Gulhane Military Medical Academy and School of Medicine, Kecioren, Ankara, Turkey.

PBRM1 identified as driver mutation

TET2 and ASXL1 were not detected

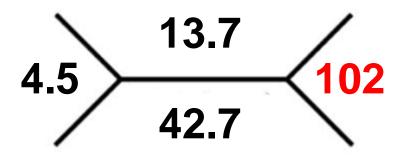
Emphasizes the need for using a robust panel of IHC for BPDCNs



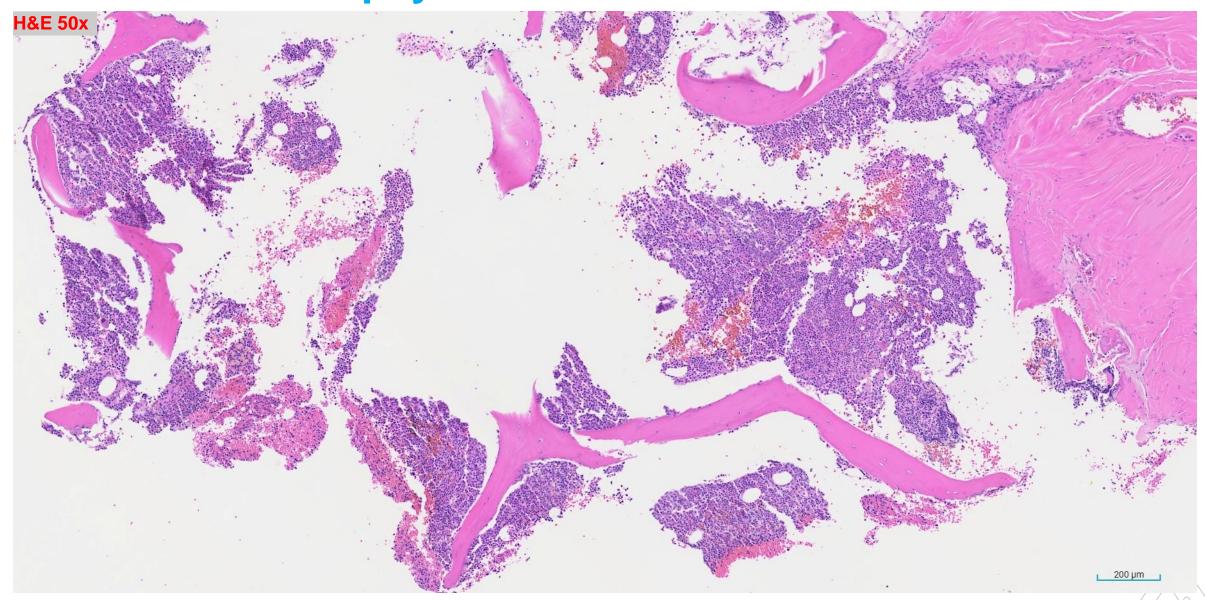
Yao, Xiaosai, and Bin Tean Teh. NATURE CELL BIOLOGY (2023)

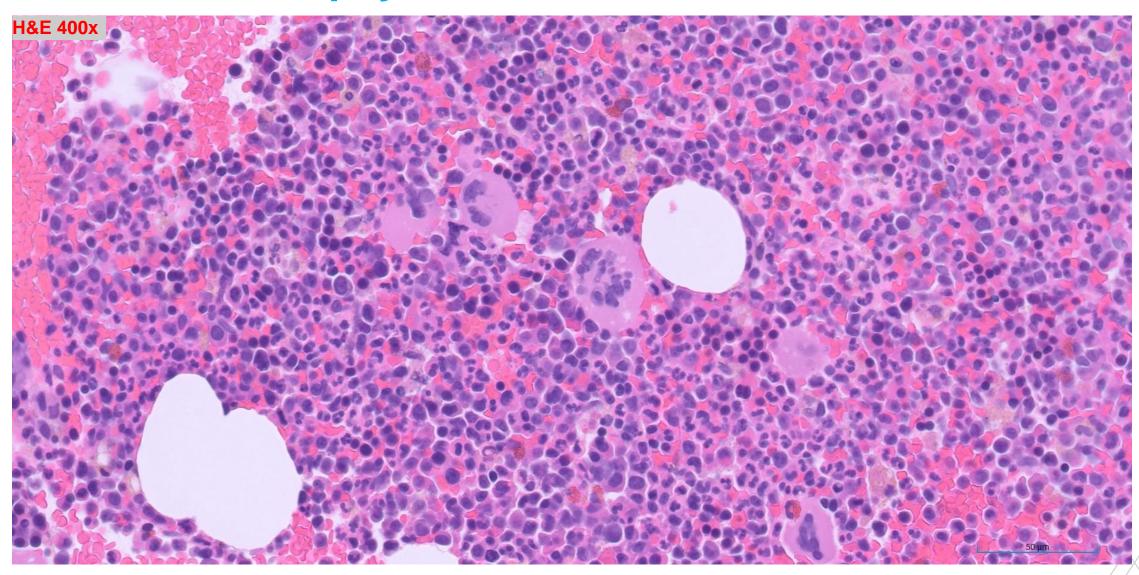
# **Clinical History**

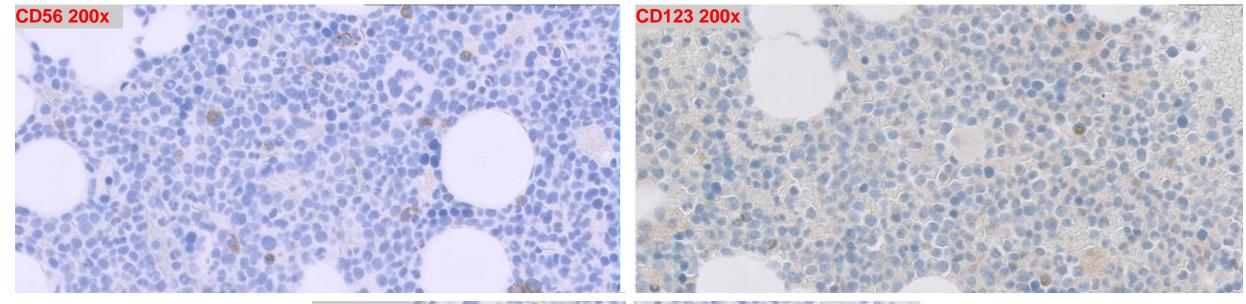
9 weeks after skin biopsy underwent bone marrow biopsy PET/CT with hypermetabolic activity of the bone

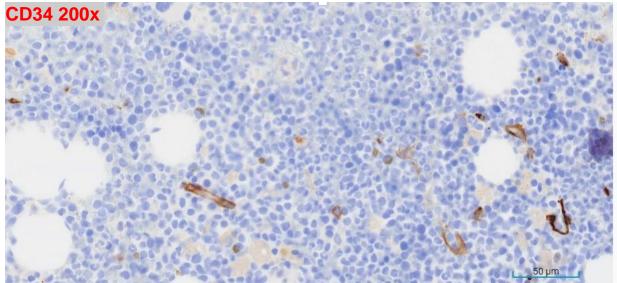


Neutrophil	31.8%
Lymphocyte	47.3%
Monocyte	15.4%
Eosinophils	3.8%
ABS Neutrophil	1.4 K/uL
<b>ABS Monocyte</b>	0.7 K/uL

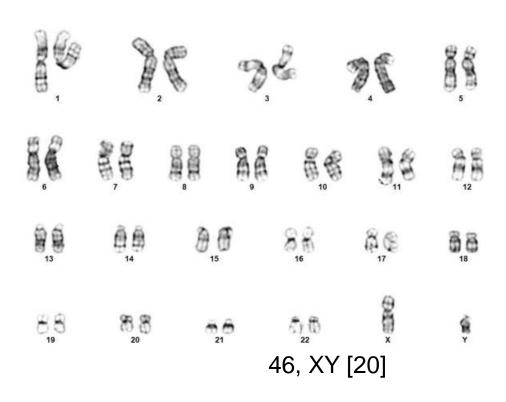








# **Cytogenetic Studies**

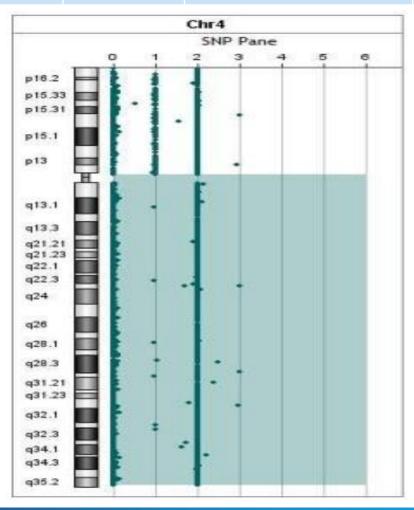


#### **FISH Panel Normal**

FISH probes tested		
KMT2A(ba)		
CFB ba		
RUNX1T1-RUNX1 DF		
BCR-ABL ES		
PML-RARA DF		
CEP8-D8Z2		
MYC ba		
TP53/D17Z1		

# **Microarray**

Chr	Cytoband	Size (kb)	Genomic Status	Gene
4	p11 – q35.2	141,543	LOH	TET2



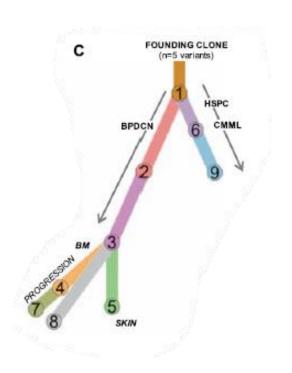
### **Bone Marrow NGS Studies**

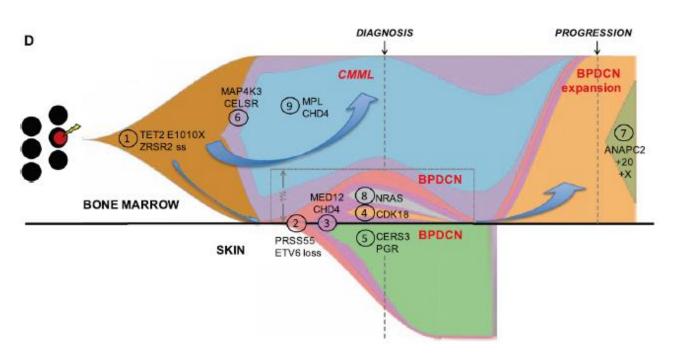
Gene	Mutation	VAF	
ASXL1	P.Q803*	48%	
SRSF2	p.P95L	54.3%	
TET2	SPLICE SITE c.3954+1G>T	95.7%	

#### **DIAGNOSIS:**

- Consistent with chronic myelomonocytic leukemia

### **BPDCN** co-occurrence with CMML, shared mutations





- BPDCN and CMML shared clonal TET2 mutations in several studies.
- Other shared mutations-

SRSF2 (Brunetti et al, Leukemia. 2017, Patnaik et al. Blood Cancer J. 2018)

### Conclusion

- CD4 negative BPDCN may potentially be a distinct entity.
- A robust panel of immunohistochemistry is useful to differentiate BPDCN from other pDC proliferations
- BPDCNs and CMML share common mutations and support a clonal relationship.



Hematopathology

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#### References

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### **CMML** Criteria

#### WHO 5th Edition

#### **Essential criteria**

- 1. Persistent absolute (≥ 0.5 x 10<sup>9</sup>/L) and relative (≥ 10%) peripheral blood monocytosis
- 2. Blasts constitute < 20% of the cells in the peripheral blood and bone marrow
- Not meeting diagnostic criteria of chronic myeloid leukaemia or other myeloproliferative neoplasms
- 4. Not meeting diagnostic criteria of myeloid/lymphoid neoplasms with eosinophilia and tyrosine kinase gene fusions (e.g. PDGFRA, PDGFRB, FGFR1, or JAK2)

#### Desirable criteria

- Dysplasia involving ≥ 1 myeloid lineages
- 2. Acquired clonal cytogenetic or molecular abnormality
- 3. Abnormal partitioning of peripheral blood monocyte subsets

#### Requirements for diagnosis

- · Essential criteria must be present in all cases
- · If monocytosis is  $\ge 1 \times 10^9$ /L: one or more desirable criteria must be met
- If monocytosis is  $< 1 \times 10^9$ /L: desirable criteria 1 and 2 must be met

#### Subtyping criteria

- Myelodysplastic CMML: WBC count < 13 x 10<sup>9</sup>/L
- Myeloproliferative CMML: WBC count ≥ 13 x 10<sup>9</sup>/L

#### **ICC 2022**

Monocytosis defined as monocytes ≥ 0.5 × 10<sup>9</sup>/L and ≥ 10% of the WBC

Cytopenia (thresholds same as MDS)

Blasts (including promonocytes) < 20% of the cells in blood and bone marrow

Presence of clonality: abnormal cytogenetics and/or presence of at least one myeloid neoplasm associated mutation of at least 10% allele frequency<sup>1</sup>

In cases without evidence of clonality, monocytes ≥ 1.0 × 10<sup>9</sup>/L and > 10% of the WBC, and increased blasts (including promonocytes), or morphologic dysplasia, or an abnormal immunophenotype consistent with CMML would be required for its diagnosis.

Bone marrow examination with morphologic findings consistent with CMML (hypercellularity due to a myeloid proliferation often with increased monocytes), and lacking diagnostic features of acute myeloid leukemia, MPN or other conditions associated with monocytosis

No BCR::ABL1 or genetic abnormalities of myeloid/lymphoid neoplasms with eosinophilia and tyrosine kinase gene fusions